Changing our views on old ideas to promote sustainable bioremediation: Fe(III) reduction and electron donors in complete TCE dechlorination

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Overview

- A (very brief) background on me and my research and consulting work
- Scope of the problem: what is needed?
- Current dogma in CI-solvent remediation
 - Fe(III) reduction is a competitive process
 - More time and more electrons
 - Dehalococcoides are the only organisms that generate ethene
- Experimental Data
 - Acetate as the sole electron donor with sustainable usage:
 - Low concentration electron donor addition
 - Less methane generated
 - Overlapping respiratory processes
 - Concurrent Fe(III) reduction and complete dechlorination
 - DHC and Non-Dehalococcoides complete dechlorination
 - Unique microbial community associated with this strategy
- Conclusions

I am primarily an academic researcher

- Ph.D. Microbiology, UMASS Amherst
- Assistant Professor University of Illinois CEE
- Associate Professor Clemson EEES
- Kavli Fellow, National Academy of Sciences
- Scientific advisory boards:
 - Battelle Chlorinated Solvents/In Situ Remediation (Monterrey Conference)
 - UMASS/AEHS Contaminated Soils Conference
 - SURF Academic liaison
 - DuPont working group on advanced geochemistry in remediation
- Research:
 - Remediation: especially bioremediation
 - Mixed biological/abiotic reactions for contaminant transformation
 - Explosives biodegradation under Fe(III)-reducing conditions
 - Combined Fe(III) reduction and complete dechlorination
 - Sustainable remediation
 - Biofuels using unbalanced fermentation
 - Mineral recovery from wastewater
 - Advanced methane production in waste systems

I am also a consultant

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- Many academics consult on their area of expertise
- We can provide answers to questions when you (the practitioners) have reached their limit of expertise, or have just hit a brick wall in the data analyses and interpretation
- As an example my firm:
 - I used to work for GeoSyntec (2001-2004)
 - I have been consulting for 8 years (One client has been with me since 2008, and I have current projects with that firm)
 - I am now on the Tersus Scientific Advisory Board
 - I have been retained once by Carus Corp. as an "on call" bioremediation expert
 - I see my strength as bridging the academic-consulting world because I have worked in both
 - I provide specialty remediation consulting on all aspects of biological and chemical remediation, biogeochemistry, field sampling plans, remediation work plans, and data analyses and interpretation to assist my clients
 - I describe my work like the TV show "House, M.D"... people (on TV...) go to him when they have reached the end of their tether for diagnostic medicine; I do diagnostic remediation analyses and design...

The Problem Entrenched Ideas in Remediation

There is no such thing as "turn key" remediation. If you ever see or hear that, think long and hard before working with the person or the company stating it

Would you stick with a doctor who claimed "turn key" services for you or your family's health...?

Chlorinated solvent remediation



Issue: the complete reductive pathway has significant geochemical and microbiological limitations → are these real or just an artifact of limited understanding

Issues in remediation

- Only one microbial species has been identified that completely reduces cis-DCE and VC to ethene (*Dehalococcoides mccartyi*; strain specific)
 - This organism works within a limited geochemical range
- Complete dechlorination has been correlated to sulfate reduction and methanogenesis (i.e. low redox potential!) – *this is incorrect*
- Fe(III) reduction has long been considered a completely competitive process – *this is also incorrect*
- Fe(III) reduction is typically the most dominant anaerobic metabolic process in subsurface environments → by negating this process we are missing the largest pool of microbial diversity
- Acetate is the key carbon intermediate in all organic matter oxidation
 - Strategies predicated on acetate as an amendment "include" a greater diversity of organisms

Issues in remediation

- Electron donors are added at much too high a concentration
- The "more time and more electrons" culture has been pervasive in the industry
- Our data (presented here and published) demonstrate that low electron donor is as good as or better than high electron donor
 - This has implications for performance and cost; it is possible to reduce TCE to ethene faster, while saving time and money, for the site
- This generates methane → a potent greenhouse gas and an absolute waste product
- Acetate is generally considered a poor electron donor for complete dechlorination, even though it is simple to use and sustains a very diverse microbial community

TCE Contaminated Site

Connecticut Aquifer Material



TCE reduction (and daughter products) and methane generation in acetate amended sediment;

TCE reduction (and daughter products) and Fe(III) reduction in *acetate* + *Fe(III) amended* sediment



Wei and Finneran, 2011, ES&T 45: 7422-

TCE reduction (and daughter products), Fe(III) reduction, and sulfate reduction in *acetate* + *Fe(III)* + *sulfate amended* sediment



VC reduction (and ethene production) and Fe(III) reduction in *acetate* + *Fe(III) amended* sediment



Wei and Finneran, 2011, ES&T 45: 7422-

1.2 2.0 TA3 - VC and ethene TB3 - VC and ethene - VC 1.0 1.5 0.8 umol/bottle umol/bottle 0.6 1.0 0.4 0.5 0.2 0.0 0.0 Fe(II) Fe(II) % Reduced Fe(II) 60 50 % Reduced Fe(II) 40 50 30 20 40 10 Sulfate Sulfate 12 8 10 mM sulfate mM sulfate 6 ומור לוווועו) 8 6 4 2 2 0 0 0 50 100 150 200 250 300 100 200 300 400 0 500 Hours Hours

VC reduction (and ethene production), Fe(III) reduction, and sulfate reduction in *acetate* + *Fe(III)* + *sulfate amended* sediment

Wei and Finneran, 2011, ES&T 45: 7422-

Fe(III) dependent "inhibition" can be due to the form of Fe(III) used in experiments



- The TCE degradation rates and daughter product distributions were different in the incubations amended with different forms of Fe(III).
- FeNTA reduction did not inhibit TCE degradation, but stimulated TCE degradation rates.
- Different microbial community structures might develop, depending on the Fe(III) speciation and bioavailability.

Microbial communities that develop in these Fe(III) reduction/complete dechlorination environments reflect two dominant groups

| clone groups | | closest relative in NCBI BLAST search | closest known relative | % in TCE only | % in TCE + FeGel | % in TCE + FeNTA |
|---------------------|-----|---|--|------------------|---------------------|---------------------|
| Dehalococcoides | G1 | Uncultured Dehalococcoides sp . clone DcocHCBPCE_48 (FJ810793.1) (99%) | Dehalococcoides sp. GT (CP001924.1) (99%) | 24.5 | 21.3 | 12.3 |
| | G2 | Dehalococcoides sp. JN18_V4_B (EF059527.1) (99%) | | 5.3 | NA | N.A |
| Geobacter | G3 | Uncultured Geobacter.sp. clone LthB63 (AM159369.1) (99%) | Geobacter humireducens (AY187306.1) (98%) | N.A | 11.7 | 19.2 |
| | G4 | Geobacter lovleyi (AY914177.1) (98%) | | 5.3 | 9.6 | 11.6 |
| | G5 | Uncultured Geobacter sp. clone: FH-33 (AB293279.1) (94%) | Geobacter humireducens (AY187306.1) (93%) | N.A | 11.7 | 7.5 |
| | G6 | Geobacter thiogenes strain K1 (NR_028775.1) (98%) | | N.A | 2.1 | 1.4 |
| other iron-reducing | 67 | Iron-reducing enrichment clone CI-A4 clone CI-A4 (DQ676996.2) (100%) | n.a. | N.A | 4.3 | 11.6 |
| clones | G8 | Iron-reducing bacterium enrichment culture clone HN-HFO29 (FJ269093.1) (100%) | n.a. | N.A | 4.3 | 8.2 |
| | G9 | Iron-reducing enrichment clone CI-A9 clone CI-A9 (DQ677001.1) (99%) | n.a. | N.A | 7.4 | 4.1 |
| other clones | G10 | Desulfosporosinus sp. 063 (GQ214051.1) (98%) | | 2.1 | 11.7 | 7.5 |
| | G11 | Uncultured bacterium clone 127 (FJ535060.1) (93%) | n.a. | N.A | N.A | 6.2 |
| | G12 | Uncultured bacterium clone FRC-AI_600 (EF507958.1) (97%) | n.a. | 6.4 | 4.3 | 4.1 |
| | G13 | Uncultured Dechlorosoma sp., isolate ALISEMBF34R34 (FM877971.1) (99%) | n.a. | 5.3 | 5.3 | 2.1 |
| | G14 | Uncultured bacterium clone J3A10 (GU139289.1) (96%) | Desulfovibrio butyratiphilus strain BSY-C (AB303306.1) (94%) | N.A | 1.1 | 1.4 |
| | G15 | Uncultured bacterium clone AN108 (GQ859927.1) (100%) | Desulfosporosinus sp. 063 (GQ214051.1) (99%) | N.A | 3.2 | 1.4 |
| | G16 | Variovorax sp. P-59, strain P-59 (AM411933.1) (97%) | | N.A | N.A | 1.4 |
| | G17 | Uncultured bacterium clone FW2_121B (GQ263435.1) (99%) | n.a. | 11.7 | 1.1 | N.A |
| | G18 | Uncultured bacterium clone SINI1098 (HM126754.1) (99%) | Azospira sp. Cu-d-1 (EF016458.1) (99%) | 8.5 | N.A | N.A |
| | G19 | Azoarcus sp. (AF482683.1) (99%) | | 5.3 | N.A | N.A |
| | G20 | Uncultured Firmicutes bacterium clone GASP-MB3S2_C03 (EF665657.1) (94%) | n.a. | 9.6 | 1.1 | N.A |
| | G21 | Uncultured bacterium clone JH-WH18 (DQ351911.1) (98%) | n.a. | 4.3 | N.A | N.A |
| | G22 | Uncultured bacterium clone FFCH16890 (EU132251.1) (97%) | n.a. | 5.3 | N.A | N.A |
| | G23 | Uncultured bacterium clone AKIW782 (DQ129364.1) (98%) | Acetonema longum DSM 6540(T) (AJ010964.1) (89%) | 1.1 | N.A | N.A |
| | G24 | Uncultured Clostridia bacterium clone X9Ba76 (AY607206.1) (98%) | Clostridium aldrichii strain P-1 (NR_026099.2) (96%) | 5.3 | N.A | N.A |

Dehalococcoides and Geobacter species were two major groups in these incubations.

FeNTA stimulated more DIRB, including Geobacter lovleyi SZ, which can transform TCE to cis-DCE with concurrent Fe(III) reduction.

Quantitative PCR data (QPCR) demonstrate that the DHC and Fe(III)-reducers are developing together



• Both *Dehalococcoides* and *Geobacteraceae* were enriched in TCE+Fe(III) incubations.

- Fel(III) reduction did not inhibit the enrichment of *Dehalococcoides* species.
- The vcrA-carrying strains was the dominant Dehalococcoides in all the samples; the highest number was found in FeGel amended sediment.

Wei and Finneran, 2011, ES&T 45: 7422-

It is all about the dissolved molecular H₂ steady state!



Wei and Finneran, 2011, ES&T 45: 7422-

Days

Perhaps some fraction of the chlorinated solvents is being mineralized to CO_2 by Fe(III) reducers, which accounts for poor mass balances in the presence of Fe(III)



The fraction of electron equivalents going to complete dechlorination is relatively small, which argues for adding less electron donor (rather than more) depending on the site

Table 1 Distribution of electron consumption in reductive dechlorination, Fe(III) reduction and methanogenesis in enrichment culture incubations with different starting acetate concentrations

| TEAPs | A. TCE + Fe(III) + stoichiometric acetate | | B. TCE + Fe(III) + $10 \times$ Acetate | | C. VC + Fe(III) + stoichiometric acetate | | $D.VC + Fe(III) + 10 \times acetate$ | |
|---|--|---------------------------------|--|---------------------------------|--|---------------------------------|---------------------------------------|---------------------------------|
| | Distribution of e ⁻ (%) | e ⁻ use/total (%) | Distribution of e ⁻ (%) | e ⁻ use/total (%) | Distribution of e ⁻ (%) | e [–] use/total (%) | Distribution of e ⁻ (%) | e ⁻ use/total (%) |
| Dechlorination | 7.01 (0.68) | 3.53 (0.18) | 4.71 (1.06) | 0.38 (0.07) | 1.64 (0.16) | 0.88 (0.11) | 0.86 (0.15) | 0.08 (0.01) |
| Fe(III) reduction | 86.77 (3.9) | 44.04 (4.96) | 58.84 (3.28) | 4.85 (0.46) | 89.79 (2.52) | 48.04 (3.76) | 57.42 (3.49) | 5.17 (0.24) |
| Methanogenesis | 6.22 (1.07) | 3.20 (0.91) | 36.45 (2.25) | 2.99 (0.13) | 8.57 (2.62) | 4.54 (1.23) | 41.72 (3.42) | 3.77 (0.51) |
| e ⁻ used/total e ⁻ available | | 50.77 | \langle | 8.22 | | 53.46 | \langle | 9.02 |

Results are presented as percent (%) of electron equivalents theoretically available from the complete oxidation of acetate. The values are the estimated fraction of electrons recovered in each terminal electron accepting process, with the remainder theoretically remaining as acetate

TCE Contaminated Site South Carolina

Influence of electron donor concentration

The issue: site electron donor concentration was already high, but more electron donors were added

The approach: use clean water to dilute the electron donor concentration (also found massive NAPL plume during my remediation investigation that was missed for almost a decade!)

~ ⁴ years GW travel time

Upgradient edge: TCE and cis-DCE (300mg/L) High electron donor Acetate 3-4mM Dissolved Hydrogen 18nM High molecular mass electron donors added in 2011 Complete dechlorination minimal to absent Chlorinated solvents mixed with petroleum hydrocarbons Other critical site data: Mn(IV) high Fe(III) high pH controlled by Fe(III) reduction

Essentially, we need to make the upgradient area mimic the downgradient area, and to do so we are going to dilute the plume using de-aerated, recirculated water (if we need to... NAPL removal may lead to MNA) Downgradient edge: TCE, cis-DCE, and VC lower Ethene present at 80% stoichiometry of VC, so complete dechlorination more active Low electron donor Acetate 100-200µM Dissolved Hydrogen 1.8nM Not influenced by 2011 electron donor amendment We got to run some lab studies...

Streambed sediments were collected at the outflow of a TCE contaminated site (TCE, cis-DCE, and VC in the sediment)

Relatively high organic carbon content given that it is shallow sediment

| · · · | TEA | | Electron Donor | | | | |
|----------------------------------|-----------------------------|------------------------------|--|-----------------------------|-----------------------------|--|--|
| Treatment | Amount TCE (µmol/bottle) | V _{TCE,liq} (mL) | H ₂ produced from Electron Donor (µmols) | Amount Substrate (mg) | Amount Substrate (µL) | | |
| Killed | 10 | 1.23 | 0 | 0 | 0 | | |
| No amend | 10 | 1.23 | 0 | 0 | 0 | | |
| 1 x Acetate | 10 | 1.23 | 30 | 0.45 | 50 | | |
| 5 x Acetate | 10 | 1.23 | 150 | 2.25 | 250 | | |
| 10 x Acetate | 10 | 1.23 | 300 | 4.5 | 500 | | |
| 1 x "RNAS Newman Zone" | 10 | 1.23 | 30 | 1.15 | 23 | | |
| 5 x "RNAS Newman Zone" | 10 | 1.23 | 150 | 5.73 | 115 | | |
| 10 x"RNAS Newman Zone" | 10 | 1.23 | 300 | 11.45 | 229 | | |
| 1 x "EOS Concentrate 598B42" | 10 | 1.23 | 30 | 0.24 | 25 | | |
| 5 x "EOS Concentrate 598B42" | 10 | 1.23 | 150 | 1.21 | 126 | | |
| 10 x "EOS Concentrate 598B42" | 10 | 1.23 | 300 | 2.42 | 252 | | |
| 1 x "CAP 18 ME" | 10 | 1.23 | 30 | 0.59 | 13 | | |
| 5 x "CAP 18 ME" | 10 | 1.23 | 150 | 2.94 | 64 | | |
| 10 x "CAP 18 ME" | 10 | 1.23 | 300 | 5.88 | 128 | | |

Table 1: Experimental matrix for TCE in batch incubations of stream bed sediments

Unamended versus acetate amended, TCE as the primary contaminant



TCE was completely reduced without adding an electron donor; this was not an electron donor limited site

> Even the low molecular mass compound acetate actually slowed the rate and extent of complete dechlorination; previous approaches at this site used high molecular mass vegetable oil based electron donors (*which was not even fermented*...!)



Days

Newman Zone (left), EOS (middle), and CAP18 (right) amended (all at 5X-10X stoichiometry) TCE as the primary contaminant

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(µmol/bottle

£



Unamended versus acetate amended, VC as the primary contaminant



Days

Days

VC, ethene, ethan

Newman Zone (left), EOS (middle), and CAP18 (right) amended (all at 5X-10X stoichiometry) VC as the primary contaminant



Unamended with VC as the primary contaminant, stimulated Fe(III) reduction with 10mmol/g ferrihydrite and completely reduced VC to ethene



Enrichment culture developed: VC reducer with stimulated Fe(III) reduction,10mmol/g ferrihydrite



Conclusions

- It is clear that the actual biogeochemical processes tied to complete reductive dechlorination are poorly understood and more work is necessary to characterize these for basic sciences and remediation practices
- Adding low concentration electron donor (stoichiometric) actually increased the extent of complete dechlorination in aquifer material (this is very different than the standard accepted practice)
- Fe(III) was concurrently reduced with ethene production (this overturns years of speculation that it is a strictly competitive process)
- Fe(III), sulfate, and chlorinated ethenes can be simultaneously reduced under this "electron donor competitive" system
- Acetate is a good electron donor for complete dechlorination, but what is the mechanism?
 - H₂ from acetate or novel community?
 - Limited methane production (less carbon footprint to waste)
- <u>This has real implications for bioremediation</u> → lower donor cost and better kinetics/extent, less methane (i.e. more sustainable)

Thank You!



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